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Roberts, Graham ; Allen, Katie ; Ballmer-Weber, Barbara ; et al

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Identifying and managing patients at risk of severe allergic reactions to food: report from two iFAAM workshops

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Abstract

Food allergy affects a small but important number of children and adults. Much of the morbidity associated with food allergy is driven by the fear of a severe reaction, and fatalities continue to occur. Foods are the commonest cause of anaphylaxis. One of the aims of the European Union funded Integrated Approaches to Food Allergen and Allergy Risk Management (iFAAM) project was to improve the identification and management of children and adults at risk of experiencing a severe reaction. A number of interconnected studies within the project have focused on quantifying the severity of allergic reactions; the impact of food matrix, immunological factors on severity of reactions; the impact of co-factors such as medications on the severity of reactions; utilising single dose challenges to understand threshold and severity of reactions; and community studies to understand the experience of patients suffering real-life allergic reactions to food. Associated studies have examined population thresholds, and co-factors such as exercise and stress. This paper summarises two workshops focused on the severity of allergic reactions to food. It outlines the related studies being undertaken in the project indicating how they are likely to impact on our ability to identify individuals at risk of severe reactions and improve their management.

1 **Background**

2 Food allergy affects around 5% of preschool children and 2% of school children and adults (1).
3 The prevalence has increased in recent decades with a doubling in peanut allergy (2) and similar
4 increases in hospitalisations due to anaphylaxis in Europe, USA and Australia (3). Food allergy is
5 the commonest cause of anaphylaxis (4), a systemic and potentially life-threatening allergic
6 reaction (5). Anxiety surrounding the fear of a severe allergic reaction has a major impact on
7 food-allergic patients and their families. For example, impaired quality of life is seen with the
8 necessary avoidance strategies, need to carry rescue medication and the associated fear of an
9 allergic reaction (6). Food-induced allergic reactions result in many presentations to hospital each
10 year and there are, often well publicised, cases of fatal anaphylaxis (7). Unfortunately these
11 severe allergic reactions are not predictable (8), and individuals with the same food allergy can
12 have very different reactions following ingestion due to co-factors (3,9). Such co-factors include,
13 among others, exercise, viral infections and several medications, whose presence around the
14 time of reaction may be associated with the development of more severe symptoms. The risks
15 associated with having food allergy are primarily described by the risk of having a severe allergic
16 reaction. The management of patients with food allergy is also driven by risk. With increasing risk,
17 more rigorous avoidance strategies are recommended and adrenaline auto-injectors are
18 prescribed to manage any future severe allergic reactions (5).

19
20 Integrated Approaches to Food Allergen and Allergy Risk Management (iFAAM) is a European
21 Union funded FP7 project. iFAAM aims to reduce the burden of food allergy through a number of
22 integrated activities. The project has developed evidence-based approaches and tools to improve
23 both the management of allergens in food as well as the management of food allergy. Many of
24 these activities are focused on the severity of allergic reactions. This paper is a summary from
25 two workshops. Firstly an iFAAM workshop held in Rome, Italy on Saturday 15th October 2016.
26 The aims of the workshop were to consider how best to (i) identify patients at risk of severe
27 reactions and situations that may impact on severity of reaction, and (ii) manage patients at risk
28 of severe reactions, particularly around high risk situations. Secondly a ILSI Europe Food Allergy
29 Symposium held in Madrid, Spain between 18th and 20th April 2018 where the final iFAAM project
30 findings were reviewed and discussed. The paper describes the iFAAM severity activities and
31 how they may lead onto improved allergen and allergy management for consumers with food
32 allergies.

33

34 Quantifying the severity of allergic reactions

35 Allergic reactions to food vary in severity, ranging from isolated oral symptoms, to relatively mild
36 cutaneous and gastrointestinal symptoms and to potentially life-threatening respiratory and
37 cardiovascular features (5). Healthcare professionals use the severity of a reaction to guide
38 treatment for that event and to inform future management of the patients. Being able to accurately
39 assess and describe the symptoms and signs of severe allergic reactions is therefore vital. A
40 number of systems have been developed to quantify the severity of allergic reactions to different
41 agents and in various circumstances, for example: venom (10,11), food (11-18), drugs (19,20)
42 and adverse reactions to allergen immunotherapy (21,22). None are in widespread use, perhaps
43 because none is ideal. They were all developed using expert opinion, and none has been
44 validated. Major limitations of the current schemes include a lack of discrimination between non-
45 anaphylactic reactions of varying severity, lack of applicability to multiple allergen types, and they
46 are not readily translatable into clinical management. For more details, see the recent European
47 Academy of Allergy and Clinical Immunology position paper (23).

48

49 To start to address these gaps, iFAAM has developed a Food Allergy Severity Score (FASS) with
50 two linked formats, a numerical nFASS developed with mathematical modelling and an ordinal
51 oFASS developed by experts' consensus with 5 grades that can be reduced to 3 (mild-moderate-
52 severe) to facilitate rapid assessment and ease of communication within most clinical settings.
53 The two scores have undergone internal validation using the EuroPrevall outpatient clinic
54 database (24), and external validation using different data sets that cover different foods,
55 paediatric and adult patients, and the whole spectrum of severity in food allergy. A manuscript
56 covering the development and validation will be submitted for publication in 2019. Further work is
57 planned to validate the system with other allergic triggers.

58

59 The availability of an appropriately developed and validated severity scoring tool will have an
60 important impact on patient care. A harmonised approach to defining severity in different
61 populations will allow data from different studies to be directly compared. Given the relationship
62 between past and future severity of reactions, this would allow the development of better
63 prediction models to identify patients at risk of future anaphylaxis. Clinically, the oFASS scale
64 should improve communication of reaction severity between primary care, acute providers and
65 allergy specialists as well as patients. This should enable specialists to better define individuals
66 who may need to take additional precautions. The nFASS is expected to be used as an outcome

67 measure in research trials; to compare severity between populations and triggers; and to analyse
68 the impact of other factors (as discussed below) and interventions in food allergy severity.

69 70 **Impact of food matrix on severity (and threshold) of allergic reactions**

71 The food matrix can affect the uptake of nutrients in the same way that the formulation of different
72 drugs may affect their absorption (25,26). This phenomenon results from the physicochemical
73 nature of the food matrix – either the natural structure of plant and animal foods (formed of
74 organelles, cells and tissues), or the fabricated foods (such as gels, dispersions, foams and
75 emulsions). Allergens can also become entrained in the structures formed by these other food
76 components. The food matrix affects the amount of a nutrient in the gut lumen which is available,
77 after digestion, to be absorbed by the body, a property generally known as bio-accessibility
78 (25,26). Coming from the field of molecular nutrition, this concept is relevant to understanding
79 how food allergens are released from food and interact with effector cells in the body to elicit an
80 allergic reaction.

81
82 The first description of a matrix effect on clinical reactivity to foods was a repeat food challenge
83 study in four patients where the fat content of a chocolate-based challenge vehicle was increased
84 to 31.5% from 22.9 % (w/w) by the addition of a white, high melting point bakery fat (27). Three
85 patients did not experience the mild oral symptoms that generally precede a reaction and
86 consequently consumed at least a 10-fold higher dose of peanut in the higher fat challenge.
87 These observations led to the hypothesis that the allergen was less bio-accessible in the oral
88 cavity when presented in the higher fat matrix. However, it is likely that the melting point of the
89 chocolate challenge was increased from 29-32°C by the use of a high melting point (47-48°C) fat,
90 as well as altering the fat crystal structure of the chocolate. A comparison of a water continuous
91 chocolate dessert matrix and dark chocolate bars in seven patients with either peanut or hazelnut
92 allergy showed the chocolate bar (35% (w/w) fat) resulted in a higher cumulative threshold dose
93 or reaction than the dessert matrix (8% (w/w) fat) (28). Magnetic resonance imaging (MRI)
94 showed the gastric residence time for the chocolate bar was much longer than the dessert which
95 might explain the delay in development of symptoms during the challenge with the chocolate bar
96 (28). There is also evidence that presentation of allergens in a baked matrix requires a higher
97 dose to be administered to elicit a reaction. For example, cow's milk allergic patients have been
98 found to reacted at lower doses to milk in a screening challenge than they did in the up-dosing for
99 oral immunotherapy with milk in a baked muffin matrix (29). However other studies have failed to

100 show a similar matrix effect with egg and cow's milk (30,31). An impact on the threshold dose and
101 severity of reactions was also observed in a group of hazelnut allergic patients when capsules
102 were used to deliver challenge doses, rather than a complex pudding matrix (32). In this study the
103 average threshold dose was increased by 3-fold, the lowest observed adverse effect level being
104 increased by 10-fold and eight subjects experiencing no reaction even after consumption of 10g
105 of hazelnut.

106
107 Insights are being gained into the mechanisms whereby the food matrix might affect allergen bio-
108 accessibility through *in vitro* digestion studies, using either batch or dynamic models of digestion.
109 An issue remains regarding the validation of such models using human studies, either by
110 analysing the intestinal contents from ileostomy patients or from intubation studies, although the
111 latter are limited to the study of liquid meals because of the constraints of nasogastric sampling.
112 However, studies are lacking directly linking *in vivo* studies on clinical reactivity to foods, and *in*
113 *vitro* studies investigating bio-accessibility of allergens during mastication and gastro-intestinal
114 digestion, and subsequent uptake into the circulation. It is inevitable that the number of studies
115 where repeat challenges have been undertaken in the same study population are small, and
116 drawing conclusions about matrix effects in populations is difficult and may be misleading,
117 especially if differences in severity of reaction, but not threshold dose are to be defined.

118
119 Addressing these gaps is a major objective of the iFAAM project with focus being placed on
120 peanut. The differences between baked matrices (cookies) and a water continuous dessert matrix
121 have been investigated in the iFAAM project. Further studies will be required using the same
122 systematic approach to identify the common rules governing bio-accessibility of food allergens in
123 complex food matrices. Such studies will inform food allergen management plans and support
124 evidence-based risk assessment of novel foods. Such knowledge is crucial to assessing the
125 allergenic risks posed by new technologies, such as 3D printing of foods, and novel functional
126 ingredients being developed to improve nutritional quality of foods and address the grand
127 challenge of food security.

128 129 **The influence of intrinsic immunological factors on reaction severity**

130 Given the pathophysiology of IgE-mediated reactions, it seems likely that the nature of food
131 allergen-specific IgE plays a role in the severity of a reaction, since IgE is a prerequisite for a

132 reaction to happen at all. For example, the qualitative nature of the IgE-response determines
133 whether a food allergic person may react to “stable” allergens (i.e. highly abundant digestion- or
134 processing-resistant allergens) such as the storage proteins of peanuts, tree nuts and seeds or to
135 “unstable” allergens such as the PR-10 related allergens cross-reacting with the birch pollen
136 allergen Bet v 1, such as hazelnut Cor a 1.04, apple Mal d 1, or even peanut Ara h 8 (33). The
137 avidity and affinity of the specific IgE-allergen interaction has been associated with the likelihood
138 of an allergic response (34). Together with prior reaction history, this may help to predict the
139 likelihood of a future severe reaction in appropriate individuals. Looking at the specific IgE in a
140 more quantitative manner, such as relating a specific IgE-titre to severity, has generated
141 equivocal results (3).

142

143 Immunoglobulins of the IgG-isotype are not considered to play a pathogenic role in food
144 hypersensitivity and their measurement is discouraged as a diagnostic marker for food allergy
145 (5,35). It has however been speculated that non-IgE isotypes - also including IgA - may offer
146 protection from food-induced IgE-mediated reactions, in a manner similar to that believed to occur
147 during allergen-specific immunotherapy. For example, food-specific IgG4 was reported to block *in*
148 *vitro* reactions in basophils and mast cells to peanut (36). Little is known though as to whether
149 such antibodies provide real-life protection and can reduce severity of food-induced allergic
150 reactions. While this has been observed in studies of tolerance induction, there are few data to
151 indicate whether IgG isotypes are useful either for predicting outcome or severity of allergic
152 reactions at oral food challenge under medical supervision. In the iFAAM project, the prognostic
153 value of specific IgG₄ was studied in a cohort of 137 patients with challenge proven peanut
154 allergy and 25 sensitized tolerant subjects. Although IgG and IgG₄ over IgE ratios were indeed
155 found to be inversely associated with severity of reactions during challenge, they did not predict
156 challenge outcomes better than IgE alone (37).

157

158 Following the cross-linking of two or more receptor-bound IgE-molecules by allergens, effector
159 cells - primarily believed to be mast cells - become activated. Basophils may also contribute to
160 reaction severity, with evidence of both basophil activation and basophil trafficking in allergic
161 reactions due to venom and food (38). The relationship between basophil activation/trafficking and
162 reaction severity needs to be further explored. Studies of the cellular compartment of IgE-induced
163 reactions, in particular mast cells, have been hampered by the question as to which anatomical
164 site actually constitutes the target organ in food allergy: when a food allergen enters the digestive

165 tract, is it absorbed systemically and distributed to target organs such as the vascular system, the
166 lungs and the upper airways to induce local mast cell activation? Or does it activate mast cells
167 lining the digestive tract, resulting in the release of inflammatory mediators from the gut which are
168 then distributed systemically? Alternatively, allergen absorption may occur across the buccal
169 mucosa (39), resulting in rapid systemic allergen distribution independent of absorption distal to
170 the mouth: such rapid absorption may explain the observation that anaphylaxis can occur within
171 minutes of ingestion.

172

173 Human mast cells originate from hematopoietic stem cells residing in the bone marrow and bear
174 the surface markers CD34, CD117 and CD13. Mast cell progenitors leave the bone marrow and
175 travel by the circulation to the peripheral sites where they home to vascular endothelium and
176 enter peripheral tissues upon recruitment. In the tissue immature lineage mast cells fully develop
177 into mature granule-containing mast cells. The differentiation and maturation of mast cells is
178 directed by cytokines in the local microenvironment. Stem cell factor (SCF) is essential for mast
179 cell differentiation. SCF binds to CD117 (c-kit) and control development and survival of mast
180 cells. Recently published studies have used *in vitro* derived mast cells to delineate a possible
181 correlation between mast cells response to allergen and asthma or atopic status, (40,41). Similar
182 studies would be needed to elucidate whether mast cells from food allergic patients are
183 predisposed to a higher activation level thus determining the severity of reactions in food allergy.

184

185 **The impact of medications as co-factors in allergic reactions**

186 The severity of allergic reactions to foods varies significantly – even within the same individual –
187 and while differences in the exposure dose may contribute to this observation, severity is clearly
188 impacted upon by a range of other influences (“co-factors”), independent of dose. One co-factor
189 thought to be responsible for increasing the severity of reactions is medications. A knowledge of
190 the impact of specific medications on the severity of allergic reactions may allow us to provide
191 information to affected individuals to help them avoid risky situations.

192

193 **Proton-pump inhibitors**

194 Gastric enzymes, such as pepsins, require an acidic environment to function. Following treatment
195 with proton-pump inhibitors (PPI), which reduces gastric acidity to pH 4.0 or higher, pepsin
196 becomes less active (42). Theoretically, this may prevent digestion-labile proteins with an

allergenic potential from being fully degraded (43), resulting in sensitisation and the development of clinical allergy. In one study, 5 of 153 patients treated with anti-ulcer medication for three months developed specific IgE to hazelnut and clinical symptoms; three of them developed symptoms under single-blind, placebo-controlled food challenges, with either pruritus or urticaria, and the remaining two upon accidental ingestion, with OAS or urticaria (44). In another report, patients with fish allergy underwent blinded food challenges with fish extract digested with gastric enzymes at pH 2.0 and 3.0. Fish extract digested at pH 3.0 triggered reactions at 10- to 30-fold lower cumulative challenge doses than extract treated at pH 2.0, and two of four patients reacted only to the latter. These data suggest that impaired digestion might put the patients with food allergy at a higher risk of developing symptoms at lower doses and at potential risk of more severe allergic reactions (45). This effect, however, has not yet been demonstrated in a well-controlled, double-blind, placebo-controlled food challenge (DBPCFC) setting, or in a larger study cohort.

Given this, within the iFAAM project the impact of PPI on severity of food-induced allergy symptoms and minimum eliciting dose has been studied at two geographically different centres, Zurich and Madrid in a randomised double-blind placebo-controlled clinical trial (EUDRA CT nr 2015-001863-38), analysing food reactivity by DBPCFC. Patients with walnut allergy have undergone three food challenges (two active, one placebo session) with a pre-treatment with either PPI or placebo. Publication of results is expected in 2019.

217

Non-steroidal, anti-inflammatory drugs (NSAIDs)

NSAID-enhanced, food-induced, allergic reactions are mainly reported in patients with food-dependent, exercise-induced anaphylaxis (FDEIA) (46) or in Lipid Transfer Protein (LTP)-sensitized individuals from Mediterranean countries. A number of case reports or series have been published (9,47). In a French case-control study, the odds ratios for severe food-induced anaphylaxis associated with aspirin and other NSAIDs were 10.8 and 8.2 respectively (48). A third (11/34) of food-induced anaphylaxis in LTP syndrome patients were cofactor-dependent (mainly NSAID) by history in one recent study (aspirin, ibuprofen, diclofenac, metamizol) (49). In another retrospective study, 58% (43/74) of patients with co-factor-induced mainly LTP-mediated food allergy, NSAID was identified as a major co-factor (50).

228

229 In just a few cases, the effect of NSAIDs on the manifestation of food allergy has been assessed
230 *in vivo*. Matsukura (46) observed an anaphylactic reaction in two patients under combined aspirin
231 and wheat challenge and Matsuo (51) observed allergic reactions in three other patients in a
232 similar setting. Brockow *et al.* has investigated aspirin as a cofactor for WDEIA, performing gluten
233 challenges under exercise with aspirin intake but unfortunately aspirin was combined with alcohol
234 as a second and therefore confounding cofactor (52). One hypothesis for the NSAID-related
235 enhancement of food allergy is the induction of increased gastrointestinal barrier permeability by
236 affecting the function of gastric epithelial tight junctions, which may increase the absorption of
237 allergens (53). Elevated gliadin concentration in serum has been observed with NSAID intake in a
238 dose-dependent manner, even in low-dose-aspirin treatment (46,51,52,55). Another hypothesis
239 postulated is a direct NSAID effect on mast cell function (9,56) and basophils (57).

240

241 Angiotensin-converting enzyme (ACE) inhibitors and beta-blocking agents

242 Both drugs have been reported to aggravate allergic reactions (refs) although other studies would
243 disagree (58-62). In a French case-control study, the odds ratios for severe anaphylaxis with co-
244 existing beta-blocker and ACE inhibitors treatment were 6.8 and 13.0 respectively (48). Risk
245 factor analysis in 4783 patients from the anaphylaxis registry of the German-speaking countries
246 calculated a modest but significant increase of odds ratio (OR increase) for the combined use of
247 these drugs (63,64). Comparing patients (adjusted for age and sex) with severity grade I/II versus
248 grade III/IV, there was no significant effect for isolated ACE inhibitor or beta-blocker intake. There
249 was only a significant effect observed for the isolated beta-blocker intake if grades I to III versus
250 grade IV were analysed (64). These data were supported by a mouse model (63). Nevertheless,
251 more studies are needed to assess the impact of ACE inhibitors and beta-blockers on the severity
252 of allergic reactions.

253

254 **The impact of exercise and physiological stress on severity of allergic reactions**

255 Other potential co-factors are exercise and physiological stress (eg sleep deprivation). They may
256 also be responsible for some of the large variation in severity of allergic reactions in individual
257 patients. One approach to look at this is to focus on threshold doses, the smallest amount of
258 allergen required to induce an allergic reaction in individuals. Population dose-distribution curves
259 describe the proportion of a population with a specific food allergy who will react to increasing
260 doses of the allergen (Figure 1). Co-factors may be able to shift population dose-distribution
261 curves to the left, resulting in a higher proportion of reactions at a lower dose and potentially

262 increasing the severity of the reaction. Conversely, as described previously, other co-factors may
263 have the opposite effect (e.g. remind readers of one) and shift the dose-distribution curves
264 towards a less sensitive population (Figure 1). Understanding these population-based dose-
265 distribution curves, the factors which might impact upon them, and the magnitude of this effect is
266 crucial to guide the advice about when precautionary allergen labelling (PAL) should be applied to
267 food products. With better informed models, higher reference doses for governmental and
268 industry-based food allergen management programs may be acceptably safe (65).

269

270 These questions have led to the development of the Threshold Reactivity Clinical Evaluation
271 (TRACE) Study, associated with the iFAAM consortium and funded by the United Kingdom Food
272 Standards Agency. One hundred peanut-allergic adults are being challenged with peanut: an
273 initial baseline peanut challenge to establish their threshold, and then three further challenges in
274 a random order, including one after sleep deprivation, another with concurrent exercise and a
275 repeat baseline challenge. A comparison of challenges with and without sleep deprivation and
276 exercise will allow within-patient variability on threshold to be determined, as well as any
277 additional impact of sleep deprivation and exercise on the threshold for reactivity. Additionally, the
278 study offers the opportunity to also look at the impact of these co-factors on reaction severity and
279 will provide dose-distribution modelling for a UK adult population.

280

281 These data will be important in informing better allergen management in the food industry as they
282 should provide a more robust evidence base for harmonised reference doses and therefore
283 action levels (65). TRACE will report in 2019.

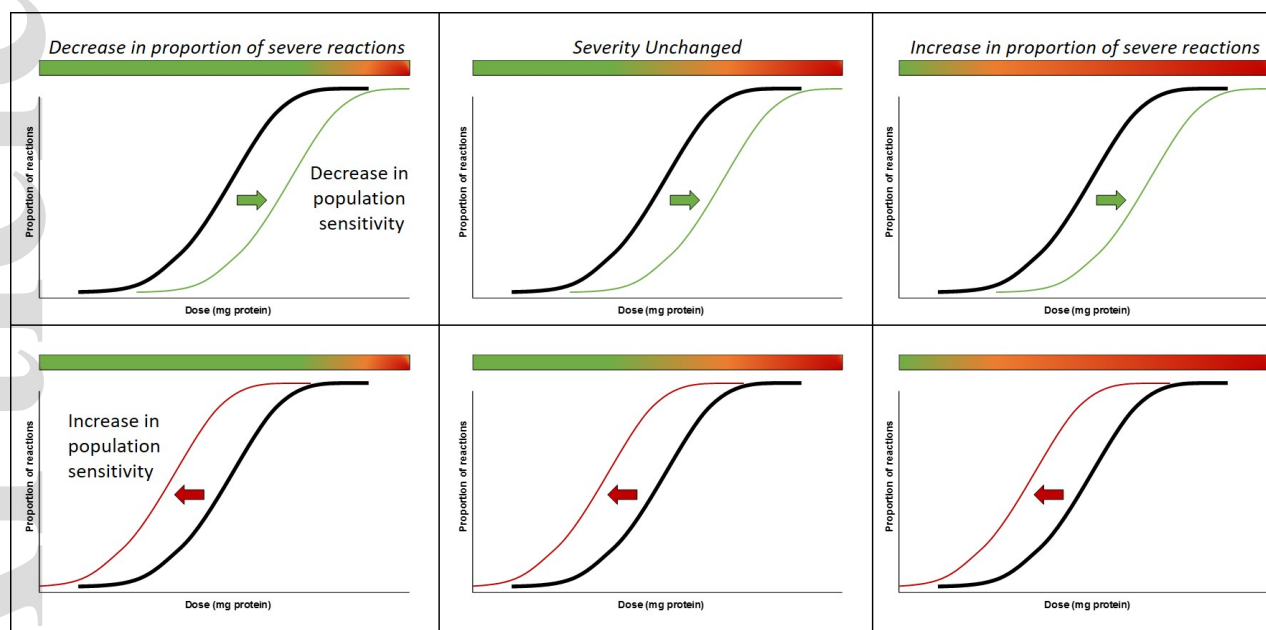
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286

287

289 **Figure 1. How do extrinsic factors affect threshold and severity of allergic reactions?**



291 Extrinsic factors may influence both the threshold and severity of allergic reactions. They may
 292 reduce the threshold dose that induces an allergic reaction in a percentage of the allergic
 293 population (bottom line) while reducing (left) or increasing (right) the severity of reactions. Other
 294 extrinsic factors may have the opposite effect increasing the threshold dose that induces an
 295 allergic reaction in a percentage of the allergic population (top line) and either reducing (left) or
 296 increasing (right) the severity of reactions. See also Hourihane and Knulst (66).

299 What can we learn from single dose challenges?

300 Population dose-distribution modelling of individual threshold doses can be used to establish
 301 public health measures such as the control of PAL. These dose-distribution curves can be used
 302 to define reference doses. For example, the dose eliciting a reaction in 1% of individuals with
 303 peanut allergy (eliciting dose, ED_{01}) has been estimated to be 0.2 mg peanut protein, using log-
 304 normal and log-logistic models (67,68). The ED_{01} was selected by the VITAL Scientific Expert
 305 Panel (VSEP) because it is predicted to protect 99% of the peanut-allergic population against any
 306 reaction. This dose can be used to inform allergen action levels as decision points to apply PAL.
 307 Alternatively a different protection goal could be chosen by using the ED_{05} , the dose at which 95%
 308 of the population are protected (69); this has the advantage of giving higher action levels
 309 involving potentially fewer products bearing PAL, albeit with more individuals remaining at risk of

allergic reactions to unintentionally present allergen(s). Whatever the reference dose used, it is clearly critical to confirm the benign nature of reactions at that dose, and particularly to ensure that their nature and frequency remain tolerable in public health terms.

313

It is important that eliciting doses derived from dose-distribution models are validated. The recently completed Peanut Allergen Threshold Study (PATS) has focused on the ED₀₅ for peanut in a single dose (66,69). Only 8/375 (2.1%) subjects had convincing objective reactions according to pre-defined criteria, all mild. In iFAAM, the same protocol has been applied to milk, egg and hazelnut to validate the ED₀₅ for each of these important allergens, albeit across much wider confidence intervals, owing to smaller numbers of participants.

320

The single dose challenge protocol has important potential clinical application. It is very easy to use and has been positively received by families, as it is much less time consuming than existing cumulative dose food challenge protocols. Food allergy-related quality of life improved in both non-reactors and reactors (66). ED₀₅ reactors could potentially be followed up by another single dose protocol using the ED₀₁ to better delineate their personal thresholds. Clinical services without the resources for full cumulative dosing food challenges could use this single dose approach safely, to identify the most dose-sensitive subjects. This could allow advice with regards to PAL to be personalised.

329

Validation of the log-normal ED₀₅ by these single dose studies, will hopefully motivate the scientific, food industry and public health communities to adopt this quantitative approach to decisions to use PAL, starting with the establishment of a defined and widely-accepted reference dose for peanut in quantitative risk assessment. This would underpin a more evidence-based use of PAL, by providing sound scientific evidence to protect the vast majority of the peanut allergic community.

336

How can community reactions help us to understand severity of allergic reactions?

Clinical reactions to food mostly occur outside the hospital or clinic setting. It is therefore important to explore severity in the community to ensure that similar patterns are seen to those observed in hospital-based studies. Such community studies are challenging, as they are inevitably observational and often retrospective, and reported severity may be prone to bias by

342 patients' personal perception and experience. The "Allergic Reactions in the Community"
343 (AllerRIC/AlleRiSC: UK, Ireland) and "Allergic Reactions" (AlleREACT: UK, Ireland, Spain,
344 Germany, Poland) studies have attempted to address some of these problems by developing and
345 validating two online reporting tools for food allergic reactions in the community, allowing for
346 reporting of incidents in near real-time (70,71). The AllerRIC/AlleRiSC study uses a multi-method,
347 longitudinal design which allows tracking of the same cohort of adult patients over a two-year
348 period (2014-2016). Adult patients with confirmed food allergies were asked to report their
349 reactions to food regardless of perceived severity, from mild incidents to anaphylaxis. Participants'
350 previous reactions and medical history were recorded in Clinical Record Forms in order to
351 compare the prospective incidence with the retrospective records. Additionally, a measure of
352 perceived severity was included in the reporting tool to explore relationships between perception
353 of severity (from 0 "extremely mild" to 9 "extremely severe") and reported symptoms (70,71). Forty
354 food-allergic incidents were reported in Ireland and the UK between October 2014 and December
355 2016. Analyses showed a significant relationship between perceived severity, reported symptoms,
356 risk perception, and psychological impact (as measured on 5-item scale adapted from Positive
357 and Negative Affect Scale, PANAS). The related AlleREACT study has gathered real time data
358 via the internet on reactions in adults with food allergy and parents of affected children from the
359 UK, Ireland, Spain, Germany, France, and Poland. To date, over 400 reactions have been
360 reported with a wide variety of severities. Results from these studies will be available in 2019.

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362

363 **Summary and conclusions**

364 Individuals experiencing severe allergic reactions to food make up an important subgroup, they
365 experience considerable morbidity, and while fatal reactions are rare, they are also unpredictable
366 (72,73). Within the iFAAM project, a Food Allergy Severity Score (FASS) has been developed
367 and validated. This has the potential to accurately describe previous allergic reactions at both a
368 simple and more complex level. Several iFAAM studies are examining the impact of the food
369 matrix on reaction severity; this would allow us to highlight which foods are particularly high risk
370 for affected individuals. The presence of co-factors may also impact reaction severity. iFAAM has
371 investigated the effect of proton-pump inhibitors, which are commonly prescribed, on DBPCFC to
372 walnut in a two centre study. The potential impact of exercise and sleep deprivation as co-factors
373 is also being examined in the associated TRACE study. Understanding the magnitude of the
374 impact of these potential co-factors on thresholds and severity will allow us to better advise
375 patients of scenarios which might place them more at risk, and provide guidance to public health

376 authorities and the food industry on the level of unintended allergen presence in foods that are
377 likely to be safe for most individuals with food allergy. These decisions are being driven by
378 population dose-distribution curves, describing the relationship between dose of allergen and
379 likelihood of reaction; these are being validated by iFAAM using single dose challenges. This
380 approach also has the potential to improve patient experience as it is a relatively simple way of
381 identifying individuals who are likely to react to very small quantities of allergen. Finally iFAAM is
382 also attempting to understand the experience of individuals who have reactions in the community;
383 this will allow us to set the experimental studies in the context of real life and to integrate the
384 impact of biopsychosocial factors, such as physiological and psychological stress, into
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386

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409 **References**

- 410 1. Nwaru BI, Hickstein L, Panesar SS, Roberts G, Muraro A, Sheikh A on behalf of The EAACI
411 Food Allergy & Anaphylaxis Guidelines Group. Prevalence of common food allergies in Europe: a
412 systematic review and meta-analysis. *Allergy* 2014; 69: 992–1007.
- 413 2. Venter, C., Arshad, SH., Grundy, J., Pereira, B., Clayton, BC., et al. (2010), Time trends in the
414 prevalence of peanut allergy: three cohorts of children from the same geographical location in the
415 UK. *Allergy* 2010; 65: 103–108.
- 416 3. Turner PJ, Gowland MH, Sharma V, Ierodiakonou D, Harper N, Garcez T et al. Increase in
417 hospital admissions due to anaphylaxis but no increase in fatalities: an analysis of UK national
418 anaphylaxis data, 1992–2012. *J Allergy Clin Immunol* 2015;135:434–442
- 419 4. Panesar SS, Javad S, de Silva D, Nwaru BI, Hickstein L, Muraro A, et al. The epidemiology of
420 anaphylaxis in Europe: a systematic review. *Allergy* 2013;68: 1353–1361.
- 421 5. Muraro A, Werfel T, Hoffmann-Sommergruber K, Roberts G, Beyer K, Bindslev-Jensen C, et al.
422 EAACI food allergy and anaphylaxis guidelines: diagnosis and management of food allergy.
423 *Allergy*. 2014;69(8):1008-25
- 424 6. Salvilla SA, Dubois AEJ, Flokstra-de Blok BMJ, Panesar SS, Worth A, Patel S, et al. Disease-
425 specific health-related quality of life (HRQL) instruments for IgE-mediated food allergy. *Allergy*
426 2014; 69: 834-844.
- 427 7. Pumphrey RS, Gowland MH. Further fatal allergic reactions to food in the United Kingdom,
428 1999–2006. *J Allergy Clin Immunol* 2007;119:1018–1019.
- 429 8. Pettersson ME, Koppelman GH, Flokstra-de Blok BMJ, Kollen BJ, Dubois AEJ. Prediction of
430 the severity of allergic reactions to foods. *Allergy* 2018; 73; 1532-1540.
- 431 9. Wölbing F, Fischer J, Köberle M, Kaesler S, Biedermann T. About the role und underlying
432 mechanisms of cofactors in anaphylaxis. *Allergy* 2013; 68: 1085-92.
- 433 10. Mueller HL. Further experiences with severe allergic reactions to insect stings. *N Engl J Med*.
434 1959;261:374-7.
- 435 11. Reisman RE. Natural history of insect sting allergy: relationship of severity of symptoms of
436 initial sting anaphylaxis to re-sting reactions. *J Allergy Clin Immunol*. 1992;90(3 Pt 1):335-9.
- 437 12. Hourihane JO, Kilburn SA, Dean P, Warner JO. Clinical characteristics of peanut allergy. *Clin*
438 *Exp Allergy*. 1997;27(6):634-9.

- 439 13. Ewan PW, Clark AT. Long-term prospective observational study of patients with peanut and
440 nut allergy after participation in a management plan. *Lancet*. 2001;357(9250):111-5.
- 441 14. Brown SG. Clinical features and severity grading of anaphylaxis. *J Allergy Clin Immunol*.
442 2004;114(2):371-6.
- 443 15. Cianferoni A, Garrett JP, Naimi DR, Khullar K, Spergel JM. Predictive values for food
444 challenge-induced severe reactions: development of a simple food challenge score. *Isr Med*
445 *Assoc J*. 2012;14(1):24-8.
- 446 16. Sampson HA. Anaphylaxis and emergency treatment. *Pediatrics*. 2003;111(6 Pt 3):1601-8.
- 447 17. Hourihane JO'B, Grimshaw KE, Lewis SA, Briggs RA, Trewin JB, King RM, et al. Does
448 severity of low-dose, double-blind, placebo-controlled food challenges reflect severity of allergic
449 reactions to peanut in the community? *Clin Exp Allergy*. 2005;35(9):1227-33.
- 450 18. Astier C, Morisset M, Roitel O, Codreanu F, Jacquenet S, Franck P, et al. Predictive value of
451 skin prick tests using recombinant allergens for diagnosis of peanut allergy. *J Allergy Clin*
452 *Immunol*. 2006;118(1):250-6.
- 453 19. Ring J, Messmer K. Incidence and severity of anaphylactoid reactions to colloid volume
454 substitutes. *Lancet*. 1977;1(8009):466-9.
- 455 20. Ring J. Anaphylactoid reactions to intravenous solutions used for volume substitution. *Clin*
456 *Rev Allergy*. 1991;9(3-4):397-414.
- 457 21. Lockey RF, Turkeltaub PC, Olive ES, Hubbard JM, Baird-Warren IA, Bukantz SC. The
458 Hymenoptera venom study. III: Safety of venom immunotherapy. *J Allergy Clin Immunol*.
459 1990;86(5):775-80.
- 460 22. Golden DB, Kwiterovich KA, Kagey-Sobotka A, Lichtenstein LM. Discontinuing venom
461 immunotherapy: extended observations. *J Allergy Clin Immunol*. 1998 Mar;101(3):298-305.
- 462 23. Muraro A, Fernandez-Rivas M, Beyer K, Cardona V, Clark A, Eller E, et al. The urgent need
463 for a harmonized severity scoring system for acute allergic reactions. *Allergy*, in press.
- 464 24. Fernández-Rivas M, Barreales L, Mackie AR, Fritsche P, Vázquez-Cortés S, Jedrzejczak-
465 Czechowicz M, et al. The EuroPrevall outpatient clinic study on food allergy: background and
466 methodology. *Allergy*. 2015;70(5):576-84.
- 467 25. Sensoy I. A review on the relationship between food structure, processing, and bioavailability.
468 *Crit Rev Food Sci Nutr* 2014;54(7):902-909.

- 469 26. Fernández-García E, Carvajal-Lérída I, Pérez-Gálvez A. In vitro bioaccessibility assessment
470 as a prediction tool of nutritional efficiency. *Nutrition Research* 2009;29(11):751-760.
- 471 27. Grimshaw KE, King RM, Nordlee JA, Hefle SL, Warner JO, Hourihane JO'B. Presentation of
472 allergen in different food preparations affects the nature of the allergic reaction--a case series.
473 *Clin Exp Allergy* 2003;33(11):1581-1585.
- 474 28. Mackie A, Knulst A, Le TM, Bures P, Salt L, Mills EN, et al. High fat food increases gastric
475 residence and thus thresholds for objective symptoms in allergic patients. *Mol Nutr Food Res*
476 2012;56(11):1708-1714.
- 477 29. Goldberg MR, Nachshon L, Appel MY, Elizur A, Levy MB, Eisenberg E, et al. Efficacy of
478 baked milk oral immunotherapy in baked milk-reactive allergic patients. *J Allergy Clin Immunol*
479 2015;136(6):1601-1606.
- 480 30 Libbers L, Flokstra-de Blok BMJ, Vlieg-Boerstra BJ, van der Heide S, van der Meulen GN,
481 Kukler J, et al. No matrix effect in double-blind, placebo-controlled egg challenges in egg allergic
482 children *Clinical & Experimental Allergy*,2013; (43) 1067–1070.
- 483 31. Remington BC, Westerhout J, Campbell DE, Turner PJ. Minimal impact of extensive heating
484 of hen's egg and cow's milk in a food matrix on threshold dose-distribution curves. *Allergy*
485 2017;72:1816–9.
- 486 32. Worm M, Hompes S, Fiedler EM, Illner AK, Zuberbier T, Vieths S. Impact of native, heat-
487 processed and encapsulated hazelnuts on the allergic response in hazelnut-allergic patients.
488 *Clinical and Experimental Allergy* 2009;39(1):159-166.
- 489 33. Asarnoj A, Nilsson C, Lidholm J, Glaumann S, Östblom E, Hedlin G, van Hage M, Lilja G,
490 Wickman M. Peanut component Ara h 8 sensitization and tolerance to peanut. *J Allergy Clin*
491 *Immunol.* 2012;130(2):468-72.
- 492 34. F. Husslik, J. Nürnberg, C. Seutter von Loetzen, T. Mews, B. K. Ballmer - Weber, J. Kleine
493 - Tebbe, et al. The conformational IgE epitope profile of soya bean allergen Gly m 4. *Clin Exp*
494 *Allergy* 2016; 46: 1484-1497.
- 495 35. Sampson HA, Aceves S, Bock SA, James J, Jones S, Lang D, et al. Food allergy: a practice
496 parameter update-2014. *J Allergy Clin Immunol.* 2014;134(5):1016-25.
- 497 36. Santos AF, James LK, Bahnson HT, Shamji MH, Couto-Francisco NC, Islam S, Houghton S,
498 Clark AT, Stephens A, Turcanu V, Durham SR, Gould HJ, Lack G. IgG4 inhibits peanut-induced

499 basophil and mast cell activation in peanut-tolerant children sensitized to peanut major allergens.
 500 J Allergy Clin Immunol. 2015;135(5):1249-56.
 501 37. M.R. Datema, E. Eller, A.H. Zwinderman, L.K. Poulsen, S.A.Versteeg, R. van Ree, Carsten
 502 Bindslev-Jensen. Ratios of specific IgG4 over IgE antibodies do not improve prediction of peanut
 503 allergy nor of its severity compared to specific IgE alone. Clin Exp Allergy 2018 in press.
 504 38. Korosec P, Turner PJ, Silar M, Kopac P, Kosnik M, Gibbs MF, et al. Basophils, high-affinity
 505 IgE receptors, and CCL2 in human anaphylaxis. J Allergy Clin Immunol 2017; 140: 750-758.
 506 39. Dirks CG, Pedersen MH, Platzer MH, Bindslev-Jensen C, Skov PS, Poulsen LK. Does
 507 absorption across the buccal mucosa explain early onset of food-induced allergic systemic
 508 reactions? J Allergy Clin Immunol. 2005;115(6):1321-3.
 509 40. Krohn,I.K. et al. Cultured mast cells from asthmatic patients and controls respond with similar
 510 sensitivity to recombinant Der P2 induced, IgE-mediated activation. Scand. J. Immunol.(2013).
 511 41. Kuehn,H.S., Jung,M.Y., Beaven,M.A., Metcalfe,D.D., & Gilfillan,A.M. Distinct PGE2-responder
 512 and non-responder phenotypes in human mast cell populations: "all or nothing" enhancement of
 513 antigen-dependent mediator release. Immunol. Lett.2011; 141: 45-54.
 514 42. Miner PB. Physiologic and clinical effects of proton pump inhibitors on non-acidic and acidic
 515 gastro-oesophageal reflux. Alimentary Pharmacology & Therapeutics 2006; 23: 25–32.
 516 43. Pali-Schöll I, Jensen-Jarolim E. Anti-acid medication as a risk factor for food allergy. Allergy.
 517 2011; 66(4):469-77.
 518 44. Schöll I, Untersmayr E, Bakos N, Roth-Walter F, Gleiss A, Boltz-Nitulescu G, et al. Antiulcer
 519 drugs promote oral sensitization and hypersensitivity to hazelnut allergens in BALB/c mice and
 520 humans. Am J Clin Nutr. 2005; 81(1):154-60.
 521 45. Untersmayr E, Vestergaard H, Malling HJ, Jensen LB, Platzer MH, Boltz-Nitulescu G, et al.
 522 Incomplete digestion of codfish represents a risk factor for anaphylaxis in patients with allergy. J
 523 Allergy Clin Immunol. 2007;119(3):711-7.
 524 46. Matsukura S, Aihara M, Sugawara M, Kunimi Y, Matsuki M, Inoue Y, Kambara T, Ikezawa Z.
 525 Two cases of wheat-dependent anaphylaxis induced by aspirin administration but not by exercise.
 526 Clin Exp Dermatol. 2010; 35(3):233-7.
 527 47. Cant AJ, Gibson P, Dancy M. food hypersensitivity made life threatening by ingestion of
 528 aspirin. Br Med J 1984; 288: 755-756.

- 529 48. Moneret-Vautrin DA, Lata arche C. Drugs as risk factors of food anaphylaxis in adults: a case-
530 control-study. *Bull Acad Natl Med.* 2009;193:351-62
- 531 49. Pascal M, Muñoz-Cano R, Reina Z, Palacín A, Vilella R, Picado C, et al. Lipid transfer protein
532 syndrome: clinical pattern, cofactor effect and profile of molecular sensitization to plant-foods and
533 pollens. *Clin Exp Allergy.* 2012;42(10):1529- 39.
- 534 50. Cardona V, Luengo O, Garriga T, Labrador-Horrillo M, Sala-Cunill A, Izquierdo A, Soto L,
535 Guilarte M. Cofactor- enhanced food allergy. *Allergy.* 2012;67(10):1316-8.
- 536 51. Matsuo H, Morimoto K, Akaki T, Kaneko S, Kusata ke K, Kuroda T, et al. Exercise and aspirin
537 increase levels of circulating gliadin peptides in patients with wheat-dependent exercise induced
538 anaphylaxis. *Clin Exp Allergy.* 2005;35(4):461-6.
- 539 52. Brockow K, Kneissl D, Valentini L, Zelger O, Grosber M, Kugler C, et al. Using a gluten oral
540 food challenge protocol to improve diagnosis of wheat-dependent exercise induced anaphylaxis.
541 *J Allergy Clin Immunol.* 2015;135(4):977-84.
- 542 53. Oshima T, Miwa H, Joh T. Aspirin induces gastric epithelial barrier dysfunction by activating
543 p38 MAPK via claudin-7. *Am J Physiol Cell Physiol.* 2008;295(3):C800-6
- 544 54. Matsuo H, Kaneko S, Tsujino Y, Honda S, Kohno K, Takahashi H, et al. Effects of non-
545 steroidal anti-inflammatory drugs (NSAIDs) on serum allergen levels after wheat ingestion.
546 *Dermatol Sci.* 2009;53(3):241-3.
- 547 55. Fujii H, Kambe N, Fujisawa A, Kohno K, Morita E, Miyachi Y. Food-dependent exercise-
548 induced anaphylaxis induced by low dose aspirin therapy. *Allergol Int.* 2008;57(1):97-8.
- 549 56. Mortaz E, Redegeld FA, Nijkamp FP, Engels F. Dual effects of acetylsalicylic acid on mast
550 cell degranulation, expression of cyclooxygenase-2 and release of pro-inflammatory cytokines.
551 *Biochem Pharmacol.* 2005;69(7):1049-57.
- 552 57. Pascal M, Muñoz-Cano R, Milà J, Sanz ML, Diaz-Perales A, Sánchez-López J, et al.
553 Nonsteroidal anti-inflammatory drugs enhance IgE-mediated activation of human basophils in
554 patients with food anaphylaxis dependent on and independent of nonsteroidal anti-inflammatory
555 drugs. *Clin Exp Allergy.* 2016;46(8):1111-9.
- 556 58. Ruëff F, Przybilla B, Biló MB, Müller U, Scheipl F, Aberer W, et al. Predictors of side effects
557 during the build up phase of venom immunotherapy for Hymenoptera venom allergy: the
558 importance of baseline serum tryptase. *J Allergy Clin Immunol.* 2010;126(1):105-11

- 559 59. Ruëff F, Przybilla B, Biló MB, Müller U, Scheipl F, Aberer W, et al. Predictors of severe
560 systemic anaphylactic reactions in patients with Hymenoptera venom allergy: importance of
561 baseline serum tryptase-a study of the European Academy of Allergology and Clinical
562 Immunology Interest Group on Insect Venom Hypersensitivity. *J Allergy Clin Immunol.*
563 2009;124(5):1047-54.
- 564 60. Stoevesandt J, Hain J, Kerstan A, Trautmann A. Over- and underestimated parameters in
565 severe Hymenoptera venom-induced anaphylaxis: cardiovascular medication and absence of
566 urticaria/angioedema. *J Allergy Clin Immunol.* 2012;130(3):698-704
- 567 61. Stoevesandt J, Hosp C, Kerstan A, Trautmann A. Hymenoptera venom immunotherapy while
568 maintaining cardiovascular medication: safe and effective. *Ann Allergy Asthma Immunol.*
569 2015;114(5):411-6.
- 570 62. Summers CW, Pumphrey RS, Woods CN, McDowell G, Pemberton PW, Arkwright PD.
571 Factors predicting anaphylaxis to peanuts and tree nuts in patients referred to a specialist center.
572 *Journal of Allergy and Clinical Immunology.* 2008; 121(3):632-e2.
- 573 63. Nassiri M, Babina M, Dölle S, Edenharter G, Ruëff F, Worm M. Ramipril and metoprolol intake
574 aggravate human and murine anaphylaxis: Evidence for direct mast cell priming. *J Allergy Clinical*
575 *Immunol* 2015; 135: 491-499.
- 576 64. Worm M, Francuzik W, Renaudin JM, Bilo MB, Cardona V, Scherer Hofmeier K, Köhli A,
577 Bauer A, Christoff G, Cichocka - Jarosz E, Hawranek T. Factors increasing the risk for a severe
578 reaction in anaphylaxis: An analysis of data from The European Anaphylaxis Registry. *Allergy.*
579 2018; 73: 1322-1330.
- 580 65. DunnGalvin A, Chan C-H, Crevel R, Grimshaw K, Poms R, Schnadt S, et al. Precautionary
581 allergen labelling: perspectives from key stakeholder groups. *Allergy* 2015; 70: 1039-51.
- 582 66. JO'B Hourihane, AC Knulst. Thresholds of allergenic proteins in foods. *Toxicology Applied*
583 *Pharmacology.* 2005;207(2 Suppl):152-6.
- 584 67. Taylor, S.L., Baumert, J.L., Kruizinga, A.G., Remington, B.C., Crevel, R.W.R., & Brooke-
585 Taylor, S. (2014) *Food Chem.Toxicol.* 63, 9–17. doi:10.1016/j.fct.2013.10.032
- 586 68. Allen KJ, Remington BC, Baumert JL, Crevel RWR, Houben GF, Brooke-Tayler S, Kruizinga
587 AG, Taylor SL (2014) Allergen reference doses for precautionary labeling (VITAL 2.0): clinical
588 implications. *Journal of Allergy and Clinical Immunology* 133:156-64

- 589 69. Hourihane JO'B, Allen KJ, Shreffler WG, Dunngalvin G, Nordlee JA, Dunngalvin A, et al.
590 Peanut Allergen Threshold Study (PATS): Novel single-dose oral food challenge study to validate
591 eliciting doses in peanut allergic children. *J Allergy Clin Immunol*, 2017; 139: 1583-1590.
- 592 70. Munro C, Semic-Jusufagic A, Pyrz K, Couch P, Dunn-Galvin A, Peek N, et al. An eHealth
593 Approach to Reporting Allergic Reactions to Food and Closing the Knowledge Gap. *Studies in*
594 *health technology and informatics*. 2015;216:320-4.
- 595 71. Pyrz KS-J, A.; Munro, Ch.; Couch, P.; Mills, C.; Hourihane, J.; Dunn Galvin, A. Developing
596 and validating a novel questionnaire to capture bio-psycho-social variables of allergic reactions in
597 the community: the AlleRiC study and the preliminary analyses. *Clinical and Translational Allergy*
598 2015; 5(Suppl 3):P1.
- 599 72. Turner PJ, Baumert JL, Beyer K, Boyle RJ, Chan CH, Clark AT, et al. Can we identify patients
600 at risk of life-threatening allergic reactions to food? *Allergy*. 2016 Sep;71(9):1241-55.
- 601 73. Hanna HJ, Emmanuel J, Naim S, Umasunthar T, Boyle RJ. Community healthcare
602 professionals overestimate the risk of fatal anaphylaxis for food allergic children. *Clin Exp Allergy*.
603 2016;46(12):1588-1595.